PCT WORLD INTELLECTUAL PROPERTY ORGANIZATION
International Bureau

INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 6 : C12N 9/14 — A2

(11) International Publication Number: WO 96/32471

(43) International Publication Date: 17 October 1996 (17.10.96)

(21) International Application Number: PCT/CA96/00223

(22) International Filing Date: 10 April 1996 (10.04.96)

(30) Priority Data:
08/419,204 10 April 1995 (10.04.95) US

(71) Applicant *(for all designated States except US)*: UNIVERSITE DE SHERBROOKE [CA/CA]; Boulevard de l'Université, Sherbrooke, Québec J1K 2R1 (CA).

(72) Inventors; and

(75) Inventors/Applicants *(for US only)*: BEAUDOIN, Adrien, R. [CA/CA]; 748 Boulevard des Vétérans, Rock Forest, Québec J1N 1Z7 (CA). SEVIGNY, Jean [CA/CA]; 144 St. Louis Street, East Angus, Québec J0B 1R0 (CA).

(74) Agents: DUBUC, Jean et al.; Goudreau Gage Dubuc & Martineau Walker, 800 The Stock Exchange Tower, Victoria Square, P.O. Box 242, Montreal, Quebec H4Z 1E9 (CA).

(81) Designated States: AL, AM, AT, AU, AZ, BB, BG, BR, BY, CA, CH, CN, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IS, JP, KE, KG, KP, KR, KZ, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TR, TT, UA, UG, US, UZ, VN, ARIPO patent (KE, LS, MW, SD, SZ, UG), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).

Published
Without international search report and to be republished upon receipt of that report.

(54) Title: ATP-DIPHOSPHOHYDROLASES, PROCESS OF PURIFICATION THEREOF AND PROCESS OF PRODUCING THEREOF BY RECOMBINANT TECHNOLOGY

(57) Abstract

The present invention relates to two ATP-diphosphohydrolases (ATPDase enzymes) isolated from bovine aorta and pig pancreas, which enzymes have a molecular weight for their catalytic unit of about 78 and 54 Kilodaltons, respectively. A first process for obtaining a highly purified ATPDase is also an object of the present invention. This process has been successfully applied to the purification of both the pancreatic and the aorta enzymes and is deemed to work in the purification of any ATPDase. For both sources of enzymes, the process allows the specific activity of the enzyme to be increased by at least 10,000 fold when compared to the activity retrieved in the crude cell homogenates. The novel process involves an ion exchange chromatography step, a separation on an affinity column, followed by an electrophoresis under non-denaturing conditions. The two enzymes purified by this process (aortic and pancreatic) are glycosylated and, when deglycosylated, have molecular weights shifted to about 56 and 35 Kdaltons, respectively. Partial amino acid sequences have been obtained for each enzyme. The partial sequences appear highly homologous with a human lymphoid cell activation antigen named CD39. An antibody directed against the porcine pancreatic enzyme cross-reacts with a protein present in endothelial cell lines and in bovine aorta (78 KDa). The high degree of homology of the pancreatic and aortic enzymes with CD39 and their cross-reactivity are indications that both enzymes are related. The pancreatic enzyme completely lacks the first 200 amino acids of CD39, which means the ATPDase activity is comprised between residues 200 and 510 of CD39. Since this is the first time that a sequence is assigned to ATPDases, a second new process for producing ATPDases by recombinant technology can also be used. Therefore, a second new process for producing an ATPDase using the CD39-encoding nucleic acid or part or variant thereof is also described.

FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AM	Armenia
AT	Austria
AU	Australia
BB	Barbados
BE	Belgium
BF	Burkina Faso
BG	Bulgaria
BJ	Benin
BR	Brazil
BY	Belarus
CA	Canada
CF	Central African Republic
CG	Congo
CH	Switzerland
CI	Côte d'Ivoire
CM	Cameroon
CN	China
CS	Czechoslovakia
CZ	Czech Republic
DE	Germany
DK	Denmark
EE	Estonia
ES	Spain
FI	Finland
FR	France
GA	Gabon
GB	United Kingdom
GE	Georgia
GN	Guinea
GR	Greece
HU	Hungary
IE	Ireland
IT	Italy
JP	Japan
KE	Kenya
KG	Kyrgystan
KP	Democratic People's Republic of Korea
KR	Republic of Korea
KZ	Kazakhstan
LI	Liechtenstein
LK	Sri Lanka
LR	Liberia
LT	Lithuania
LU	Luxembourg
LV	Latvia
MC	Monaco
MD	Republic of Moldova
MG	Madagascar
ML	Mali
MN	Mongolia
MR	Mauritania
MW	Malawi
MX	Mexico
NE	Niger
NL	Netherlands
NO	Norway
NZ	New Zealand
PL	Poland
PT	Portugal
RO	Romania
RU	Russian Federation
SD	Sudan
SE	Sweden
SG	Singapore
SI	Slovenia
SK	Slovakia
SN	Senegal
SZ	Swaziland
TD	Chad
TG	Togo
TJ	Tajikistan
TT	Trinidad and Tobago
UA	Ukraine
UG	Uganda
US	United States of America
UZ	Uzbekistan
VN	Viet Nam

TITLE OF THE INVENTION

ATP-DIPHOSPHOHYDROLASES, PROCESS OF PURIFICATION THEREOF AND PROCESS OF PRODUCING THEREOF BY RECOMBINANT TECHNOLOGY.

FIELD OF THE INVENTION

The present invention relates to a process of purification to homogeneity of ATP-diphosphohydrolases involved in numerous nucleotide and nucleoside receptor-mediated physiological functions, namely platelet aggregation, vascular tone, secretory, inflammatory and excretory functions and neurotransmission. These enzymes, which have been particularly obtained from bovine aorta and pig pancreas have been purified and their catalytic unit identified. The partial amino acid sequences of each ATPDase show a high degree of homology with a lymphoid cell activation system named CD39.

BACKGROUND OF THE INVENTION

ATP-diphosphohydrolases (ATPDases) or apyrases (EC 3.6.1.5) have been found in plants, invertebrates and vertebrates. The enzyme catalyses the sequential hydrolysis of the γ- and β-phosphate residues of triphospho- and diphosphonucleosides. These enzymes are generally activated in the presence of divalent cations Ca^{+2} or Mg^{+2} and inhibited by sodium azide. In plants, the enzymes are found in the cytoplasm, in soluble or membrane-associated forms, and are generally more active at acid pH. Their precise function is not known, but there is some evidence that they are involved in the synthesis of carbohydrates. In invertebrates, the enzymes are more active at neutral or alkaline pH. Found mainly in saliva and in salivary glands of hematophagous insects, an antihemostatic role has been demonstrated. In vertebrates, a limited number of studies have already defined a diversity of ATPDases. The catalytic site of these enzymes is generally exposed to extracytoplasmic spaces (ectoenzymes). By their

location and kinetic properties, these different types of ATPDases could influence the main systems of the organism, namely vascular and nervous systems. Their specific role in these systems is determined by the presence of purine and pyrimidine receptors which react with triphosphonucleosides and their derivatives at the surface of numerous cell types.

Presence of both ectoATPase and ectoADPase activities in the vascular system has been known for many years, and up until the work of Yagi et al. (1989), they were attributed to two distinct enzymes. The latter purified these activities and showed that in bovine aorta, a single enzyme was responsible for the sequential hydrolysis of ATP and ADP. A mammalian ATPDase had been first described in the pancreas (Lebel et al., 1980) and was further reported in several other tissues. Yagi et al. (1989) proposed that the enzyme from aorta was similar to the previously reported mammalian ATPDase from pancreas and that it was associated with the intima of bovine aorta.

Purification to homogeneity was demonstrated by SDS-polyacrylamide gel electrophoresis (PAGE) and silver staining. The apparent molecular weight of the pure enzyme was estimated at 110 KDa. The existence of the ATPDase in the bovine aorta was corroborated by Côté et al. (1991) who, by showing that identical heat and irradiation-inactivation curves with ATP and ADP as substrates, assigned to the same catalytic site the ATPase and ADPase activities. A comparison of the biochemical properties led Côté et al. *supra* to propose that the bovine aorta enzyme was different from the pancreas ATPDase. Indeed, the enzymes have different native molecular weights, optimum pH and sensitivities to inhibitors. They proposed to identify pancreas enzyme as type I and the aorta enzyme as type II. In the bovine aorta, the enzyme was found to be associated with smooth muscle cells and endothelial cells and could inhibit ADP-induced platelet aggregation. Côté et al. (1991) further showed that concurrent addition of ATPDase and ATP to platelet-rich plasma resulted in an

immediate dose -dependent platelet aggregation caused by the accumulation of ADP, followed by a slow desaggregation attributable to its hydrolysis to AMP. In the absence of ATPDase, ATP did not induce any aggregation while ADP initiate an irreversible aggregation which extent is limited by the ADPase activity of the enzyme. ATPDase also attenuated the aggregation elicited by thrombin and collagen but not by PAF (Platelet Activating Factor), the first two agonists having an effect mediated by platelet ADP release. It was therefore suggested that ATPDase had a dual role in regulating platelet activation. By converting ATP released from damaged vessel cells into ADP, the enzyme induced platelet aggregation at the sites of vascular injury. By converting ADP released from aggregated platelets and/or from hemolyzed red blood cells to AMP, the ATPDase could inhibit or reverse platelet activation, and consequently limit the growth of platelet thrombus at the site of injury. In their attempt to further characterize the aorta ATPDase, the

present inventors have developed a new process for producing highly purified ATPDases. They have established a procedure by which its specific activity can be increased over and above the activity of a crude cell preparation by more than 10000-fold. They also discover that the purified enzyme (the catalytic unit) had a molecular weight different from the one previously reported for the native form of the enzyme (190 KD by using the irradiation technique), suggesting that the enzyme may exist in a multimeric form in its native state. Partial amino acid sequences of both bovine aorta and porcine pancreatic ATPases have been obtained.

In a completely different field, Maliszenski et al. (1994) have published the sequence of a human lymphoid cell activation antigen designated CD39. Another group (Christoforidis et al. 1995) described the purification of a human placenta ATPDase of a molecular weight of 82 KDa. Its partial amino acid sequence shows a high degree of homology with CD39.

When the above mentioned partial amino acid sequences were entered in GenBank for verifying the presence of any homologous sequence, complete homology was surprisingly found for some of these fragments with the CD39 gene product. The complete sequences of the ATPDases remain to be obtained. Assuming that CD39 is an up to date unknown ATPDase, a process for producing ATPDases by recombinant technology is now possible, and CD39 can now be used to reduce platelet aggregation and thrombogenicity.

STATEMENT OF THE INVENTION

It is an object of the present invention to provide two ATPDases isolated from bovine aorta and porcine pancreas, which enzymes have a molecular weight for their catalytic unit of about 78 and 54 Kilodaltons, respectively. A novel process for obtaining a highly purified ATPDase is also an object of the present invention. This process has been successfully applied to the purification of both the pancreatic and the aorta

enzymes and is deemed to work in the purification of any ATPDase. For both sources of enzymes, the process allows the specific activity of the enzyme to be increased by at least 300 fold when compared to the activity retrieved in the microsomial fraction of these cells as previously reported for an aortic and pancreatic proteins of a native molecular weight of about 190 and 130 KDa, respectively.

The two ATPDases purified to homogeneity were partially sequenced. These sequences have shown striking similarities with a human lymphoid cell activation antigen named CD39 (Maliszenski et al., 1994). Since the molecular weight of CD39 and its glycosylation rate appears to define a human counterpart for the present bovine aortic ATPDase, it is the first time that a sequence is assigned to an ATPDase. A process of producing an ATPDase by recombinant technology is now possible using a host cell expressing the CD39 human protein, its homologous sequences in

bovine and porcine species, and variants and parts thereof.

The present invention also relates to the use of CD39 and of the above bovine and porcine homologous proteins for reducing platelet aggregation and thrombogenicity.

DESCRIPTION OF THE PRESENT INVENTION

The research team to which the present inventors belong has already characterized the pig pancreatic ATPDase, and the latter reassessed the properties of the bovine aorta enzyme. They confirmed that the aorta ATPDase was different from its pancreatic counterpart. They have found previously (Côté et al., 1992) that the aorta enzyme (isolated from a microsomal fraction of the cells) had a molecular weight of about 190 kDa in its native state. In their work for extensively purify this enzyme, they found that the highly purified enzyme had a molecular weight on SDS-PAGE of about 78 KDa. Yagi et al. (1989) have already shown that an ATPDase purified

to homogeneity had a molecular weight of 110 KDa. After purifying the enzyme by the present method, the 110 kDa band was indeed absent from SDS-PAGE. A unique band migrating of an estimated weight of 78 KDa was rather revealed. The confirmation of the identity of the purified enzyme was achieved by binding FSBA, an ATP analog binding the enzyme, to the separated and blotted enzyme. The use of anti-FSBA antibodies revealed the presence of the bound enzyme and this binding was inhibited with ATP and ADP. The same procedure was applied to confirm the identification of the pancreas ATPDase Type I.

The present process allows the purification of ATPDases to a very high level. In the aorta, the purified enzyme has a specific activity which is increased by at least 300 fold compared with the specific activity of microsomal fraction (already enriched by about 30 fold from the crude cell preparation).

The bovine aorta and porcine pancreatic ATPDases have been partially sequenced, and the sequences have been found to be highly homologous to a human lymphoid cell activation antigen designated CD39 (Maliszenski et al., *op. cit.*). The complete sequences of the ATPDases types I and II have not been obtained yet. If one assumes that CD39 gene product is an ATPDase type II, the present invention therefore contemplates the use of CD39 in the reduction of platelet aggregation and of thrombogenicity, as well as a process of making ATPDases using the CD39 sequence, variants or parts thereof (recombinant technology).

The present invention will be described hereinbelow with reference to the following Examples and Figures which purpose is to illustrate rather than to limit the scope of the present invention.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 illustrates the protein composition of the bovine aorta ATPDase (type II) at the different purification steps as determined by SDS-PAGE. Electrophoresis was run in a 7-12.5% polyacrylamide gel. Proteins were stained with Coomassie Blue or silver nitrate dye. MW standards: 97.4, 66.2, 45.0, 31.0, 21.5, 14.4 KDa; particulate fraction (part. fract.), 100 μg; DEAE-agarose fraction, 35 μg; Affi-Gel blue fraction, 20 μg; the lower band of activity was cut out from the non-denaturing gel (N.D. gel); sample buffer alone (Control).

Figure 2 illustrates a Western blot of FBSA labelled protein (ATPDase type II) isolated from Affi-Gel blue column. Labelled proteins were separated on a 8-13.5% gradient gel by SDS-PAGE, transferred to Immobilon-P membrane, incubated with a rabbit antibody anti-FBSA (1:10,000) and detected by a secondary antibody conjugated to alkaline phosphatase (1:6,000). Twenty μg of protein from Affi-Gel blue column fraction was used

for the assays: incubation with FBSA (FBSA); incubation with FBSA with competing Ca-ATP (FBSA+ ATP); incubation without FBSA (no FBSA). MW standards are the same as in Figure 1.

Figure 3 illustrates the SDS-PAGE protein patterns at the different steps of the purification procedure and after N-glycosidase F digestion of the Affi-Gel blue fraction. Protein samples were fractionated on a 8-13.5% polyacrylamide gradient. A) One unit of N-glycosidase F (silver nitrate stain); B) Six μg from the Affi-Gel blue fraction incubated for 12 h without N-glycosidase F (silver nitrate stain); C) Idem as B with 1 unit of N-glycosidase F (silver nitrate stain); A') Same as A (Coomassie blue stain); B') Same as B (Coomassie blue stain); C') Same as C (Coomassie blue stain); D) MW standards: 97.4, 66.2, 45.0, 31.0, 21.5, 14.4 kDa (Coomassie blue stain), E) ZGM (zymogen granule membrane), 60 μg (Coomassie blue stain); F) Active fraction from DEAE-agarose column, 25 μg (Coomassie blue stain); G) Active fraction from Affi-Gel blue column, 6

μg (Coomassie blue stain); G') Same as G (silver nitrate overstain); H) Activity band located after PAGE under non-denaturing conditions (silver nitrate overstain); I) Control, band located just above the activity band after PAGE under non-denaturing conditions (silver nitrate overstain).

Figure 4 shows a Western blot of FSBA labelled samples of the pancreatic enzyme type I fraction. Labelled sample were loaded on a 7-12% polyacrylamide SDS-gel, transferred to Immobilon-P membrane, incubated with the rabbit antibody anti-FSBA and detected by a secondary antibody conjugated to alkaline phosphatase. Six μg of Affi-Gel blue column were used in lanes B), C) and D). A) MW standards: 97.4, 66.2, 45.0, 31.0, 21.5, 14.4 kDa; B) FSBA; C) FSBA + competing ADP; D) No labelling.

Figure 5 shows a Western blot of human endothelial cell extracts labelled with an antibody directed against a fragment common to ATPDase type I and CD39. The ATPDase type II (78KDa) is clearly detected as well as low amounts of ATPDase type I (54KDa).

Example 1

PURIFICATION OF THE ATPDase type II

a) Isolation of the particulate (microsomal) fraction from the bovine aorta:

Bovine aorta, obtained from a local slaughterhouse, were kept on ice and processed within one hour after the death of the animals. All steps were carried out at 4°C. The inner layer was stripped out manually, passed through a meat grinder, and homogenized (10%) with a Polytron™ in the following solution: 95 mM NaCl, Soybean Trypsin Inhibitor (20 µg/mL), 0.1 mM Phenyl-methyl-sulphonyl-fluoride (PMSF) and 45 mM Tris-HCl pH 7.6. After filtering with cheesecloth, the homogenate was centrifuged at 600 X g for 15 minutes with a Beckman JA-14 centrifuge at 2100 RPM. The supernatant was recovered and centrifuged at 22,000 X g for 90 minutes with the same centrifuge at 12,000 RPM. The resulting pellet was suspended in 0.1 mM PMSF and 1 mM $NaHCO_3$ pH 10.0 with a Potter Elvejehm™ homogenizer at a dilution of 3 to 6 mg of protein per mL. The suspension was

loaded on a 40% sucrose cushion and centrifuged at 100,000 X g for 140 minutes with a SW 28 Beckman rotor. The enzyme was recovered on the cushion and kept at 4°C overnight. This membrane preparation was then suspended in 12 volumes of 0.1 mM PMSF and 1 mM $NaHCO_3$ pH 10.0 and centrifuged at 240,000 X g for 45 minutes in a SW 50.2 Beckman rotor. The pellet was rinsed twice: once with o.1 mM PMSF and 30 mM Tris-HCl ph 8.0 and once with 2 mM EDTA and 30 mM Tris-HCl pH 8.0. The final pellet was suspended in 7.5% glycerin and 5 mM Tris-HCl ph 8.0 at a concentration > 1 mg of protein per mL and frozen at -20°C, or directly solubilized. At this stage, the specific activity of the ATPDase was enriched by about 33 fold.

b) Solubilization and column chromatographies:

The particulate fraction (pf) was solubilized with 0.3% Triton X-100™ and 30 mM Tris-HCl pH 8.0 at a concentration of 1 mg/mL protein and centrifuged at 100,000 X g for 1 hour in a SW 50.2 Beckman rotor. All further steps involving a detergent are practised with

Triton X-100, but any similar detergent (a non-ionic detergent) may be used for achieving the purpose of this invention. The supernatant was loaded on an ion exchange column, preferably containing diethylaminoethyl (DEAE), like DEAE-Bio Gel A Agarose™, preequilibrated with 0.1% Triton X-100™, 7.5% glycerin and 10 mM Tris-HCl pH 8.0. The protein was eluted in the same buffer by a NaCl gradient (0.03 to 0.12 M), followed by a 0.1% Triton X-100™ and 2 M NaCl wash. Active fractions were pooled in 0.1X buffer E (5X buffer E: 0.5% Triton X-100™, 960 mM glycine, 125 mM Tris-HCl pH 7.0) and electrodialysed in 15 mL cuvettes by an ISCO™ electro-eluter according to the following technique: 1X buffer E was loaded in the apparatus and a 15 mA current was applied per cuvette. The 1X buffer E was changed 4 times at 50 minute intervals. The dialysate was equilibrated at pH 5.9 with 200 mM histidine adjusted to pH 4.0 with HCl (about 20 mM final) and loaded on an Affi-Gel™ blue column preequilibrated with 0.07% Triton X-100™, 7.5% glycerin, 30 mM histidine and 30 mM Tris-HCl pH 5.9. Proteins

were eluted by a linear gradient from 100% buffer A to 100% buffer B (buffer A (80 ml): 0.07% Triton X-100™, 7.5% glycerin and 10 mM Tris-HCl pH 6.5; buffer B (80 ml): 1M NaCl, 0.07% Triton X-100™, 7.5% glycerin and 10 mM Tris-HCl pH 7.5), followed by a 1M NaCl, 0.1% Triton X-100™, 100 mM Tris-HCl pH 8.5 wash. The active fraction was dialysed against 0.05% Triton X-100™, 1 mM Tris-HCl pH 8.0, concentrated on a 1 ml DEAE-agarose column as described above, eluted in 0.4 M NaCl, 0.07% Triton X-100™, 10 mM Tris-HCl pH 8.0 and dialysed against distilled water.

c) Separation by polyacrylamide gel electrophoresis (PAGE) under non-denaturing conditions:

This type of gel allows for separating proteins upon their molecular weight and electrical charge while preserving their activity in such a way that this activity can be measured after migration. Two polyacrylamide preparations were poured between two glass plates to form a gradient and polymerized. The 4%

acrylamide solution was composed of: 4.5 mL of separating buffer (Tris 1.5 M pH 8.8+ 0.4% Triton X-100™), 2.5 mL acrylamide 30%, 180 µL Na deoxycholate 10%, water up to 18 mL, 60 µL APS 10% and 7 µL TEMED. The 7.5% acrylamide solution was composed of the same ingredients except for the volume of acrylamide: 4.5 mL. A stacking gel was extemporaneously prepared and poured at the top of the separating gel, the stacking gel was composed of: 2.5 mL of stacking buffer (Tris-base 0.5 M pH 6.8), 6.1 mL of water, 1.34 acrylamide 30%, 0.1 mL Na deoxycholate 10%, 0.1 mL Triton X-100™, 50 µL APS 10% and 10 µL TEMED. Wells are formed in this layer during polymerization. Two volumes of the sample obtained after DEAE-agarose or Affigel Blue columns were added to one volume of sample buffer of the following composition to obtain about 100 µg proteins: 0.07% (v/v) Triton X-100™, 1.5% (w/v) Na deoxycholate, 10% glycerol, 65 mM Tris-base and 0.005% bromophenol blue. The suspended sample was allowed to stand 10 minutes on ice and centrifuged. The supernatant was loaded on gel. The

proteins were migrated at 4°C at a 20 mAmp power in reservoir buffer (0.1% Triton X-100, 0.1% sodium deoxycholate, 192 mM glycine and 25 mM Tris pH 8.3). For revealing activity in the separated bands, the latter were placed in a dosage buffer (Tris-base 66.7 mM, imidazole 66.7 mM, $CaCl_2$ 10 mM, pH 7.5). After preliminary incubation for 30 minutes at 37°C, the substrate (ADP or ATP) 5 mM was added. After 2 to 10 minute incubation, a white calcium phosphate precipitate significative of ATP diphosphohydrolase activity is formed. Three bands are seen for the aorta enzyme and one for the pancreas (these bands were all revealed on gel by silver overstaining). For further characterization, the most active band was loaded on an SDS-PAGE according to Laemmli (1970) and a single band appeared on the gel after silver nitrate staining, which is indicative of an enzyme purification to homogeneity after the non-denaturing gel. Figure 1 shows the high sensitivity of detection conferred by the use of silver staining compared to a conventional Coomassie blue

staining (see lanes 4 and 5). The active band purified from the gel has a molecular weight of 78 KDa when migrated on SDS-PAGE.

d) ATPDase assays during chromatographic steps:

Enzyme activity was determined at 37°C in the following incubation medium: 50 mM Tris-imidazole (pH 7.5), 8 mM $CaCl_2$ and 0.2 mM substrate (ATP or ADP). Phosphorus was measured by the malachite green method according to Baykov et al. (1988). One unit of enzyme corresponds to the liberation of 1 µmol of phosphate per minute per mg of protein at 37°C. Proteins were estimated by the technique of Bradford (1976).

The ATPDase activity retrieved in isolated fractions are summarized in the following Table:

Table 1. ATPDase purification of the bovine aorta ATPDase type II

Step	Total protein mg	Total activity units	Specific activity units/mg	Yield %	Purification factor -fold	Hydrolysis rat ATP/ADP
Particulate fraction (pf)	293	263	0.9	-	(33)*	1.5
pf + Triton X-100	293	117	0.4	100	1	1.4
100,000 *g* supernatant of solubilized pf	186	91.2	0.5	78	1.2	1.3
DEAE column	15.1	72.2	4.8	62	11.9	1.1
Affi-Gel blue column	2.76	57.8	21	49	53	1.1
Con A	0.61	33.5	55	29	138	1.1

Details on the purification and condition assays are described in the disclosure. A representative out of five complete purification procedures is shown with ADP as substrate. Determinations were routinely carried out in triplicate. * The starting particulate fraction shows a 33 purification folds as compared to the homogenate (Côté 1991).

e) Confirmation of the identity of ATPDase:

The fraction eluted from Affi-gel was labelled with 5 '-p-fluorosulfonylbenzoyl adenosine (FBSA), a marker which forms covalent bonds with adenosine-binding proteins. FSBA blocked the enzyme activity and excess of ATP or of ADP prevents this effect. In addition, FSBA efficiently bound the purified enzyme, as monitored by a Western blot technique using an antibody directed to FSBA, which binding is prevented in the presence of ATP (see Figure 2) or ADP (data not shown).

The results obtained on SDS-PAGE shows that the enzyme was purified to homogeneity when using the successive steps of solubilization of the particulate fraction, first purification on an ion exchange column, second purification on an affinity column and third purification on non-denaturing electrophoretic conditions. The Affigel Blue column did not achieve purification to homogeneity but allowed a much higher recovery then the 5 ' AMP-Sepharose™ used by Yagi et al. (about 7 fold higher). Moreover, the use of the Affigel

column and the non-denaturing gel allowed us to purify an enzyme that is different from the one disclosed by Yagi.

f) ATPDases are glycosylated proteins:

Purification on Concanavalin A column:

Further purification of the Affi-Gel blue fraction of aorta enzyme was also obtained with Con A agarose column. Briefly, Con A (4 ml beads) and the protein sample from the Affi-Gel blue column were preequilibrated with 0.05 % Triton X-100, 100 mM NaCl, 1 mM $CaCl_2$, 1 mM $MnCl_2$ and 20 mM PIPES, pH 6.8, at room temperature. The protein sample was passed through the column at a flow rate of 3 ml/h, 40 ml of the preequilibration buffer was then added to wash the unbound materials at a flow rate of 10 ml/h. The activity was eluted with 20 ml of 0.5 M Me-α-D-mannopyranoside diluted in the preequilibration buffer. The purified sample was dialysed and concentrated on a mini-DEAE column as described above.

Precipitation of ATPDase activity with lectin-agarose:

Four lectins conjugated to agarose were tried: Con A, WGA, Soybean agglutinin and UEA. Experiments were carried out at room temperature for Con A, and at 4°C for the other agglutinins. One hundred µl of each 50% slurry were put in a microcentrifuge tube and washed 4 times with buffer A: 0.05% Triton X-100, 100 mM NaCl and 20 mM PIPES pH 6.8. In the case of Con A, 1 mM $CaCl_2$ and 1 mM $MnCl_2$ were added to this buffer. Twenty µg of ATPDase purified from the Affi-Gel blue column, equilibrated in buffer A, were added to the lectin-agarose beads and rocked for 45 min, then centrifuged for 1 min. The supernatant was kept and the beads were washed 3 times with 1 ml buffer A. Protein bound to the lectins was eluted with 150 µl of 500 mM of the appropriate sugar in buffer A, rocked for 30 min and centrifuged. The elution step was repeated once and the 2 eluates were pooled. The sugar used to eluate proteins from Con A, WGA, Soybean and UEA were

Me-α-D-mannopyranoside, D-GlcNAc, D-GalNAc and L-Fuc respectively.

Table 2. ATPase binding to lectins

Lectin-agarose	Fractions	Relative ADPase activity	Presence of the 78 kDa band on SDS-PAGE	Sugar specificity
Con A	Supernatant	5%	traces	Mannose,
	Bound	95%		Glucose
	Eluted	62%	+	
WGA	Supernatant	5%	traces	GlcNAc, NeuNAc,
	Bound	95%		Mannose structure §
	Eluted	69%	+	Sialic acid §
Soybean	Supernatant	100%	+	GalNAc
	Bound	0%		
	Eluted	0%	-	
UEA	Supernatant	100%	+	Fucose
	Bound	0%		
	Eluted	0%	-	

Twenty μg of ATDPase fraction purified by Affi-Gel blue chromatography were incubated separately with four lectins conjugated to agarose, centrifuged, and the supernatants were collected. Lectins-agarose beads were then washed. Bound proteins were finally eluted with the appropriate sugar as described in the disclosure. This experiment has been done twice in triplicate and the mean is presented. In parallel, the supernatant and the eluted fraction were put on SDS-PAGE, stained with silver nitrate, and looked for the presence of the 78 kDa. The sugar specificity of each agglutinin is also presented.

§ Weak affinities

Only WGA bound the ATPDase type II as for Con A. ATPDase binding to these two lectins is indicative of a specificity for the sugars glucose and/or mannose and/or GlcNAc (Glucosamine-N-Acetyl) and/or NeuNAc (Neuraminic-N-Acetyl).

The deglycosylated form had a molecular weight of about 56 KDa, which suggests that about 5 to 11 glycosyl chains are present on the 78 KDa protein (assuming that a glycosyl group may have a molecular weight of 2 to 4 KDa).

Example 2

PURIFICATION OF THE ATPDase type I

The procedure described in Example 1 has been followed for purifying the pancreatic ATPDase type I enzyme, starting from the zymogen granule membrane of pig pancreas.

In deglycosylation experiments, the molecular weight of the catalytic unit has been shown to be shifted from 54 to 35 KDa. Therefore, the chemical

procedure exemplified above is deemed to apply to the purification of ATPDases in general.

h) Level of enrichment:

The level of enrichment is determined from the data shown in Table 1 for aorta ATPDase type II and from the following Table 3 obtained for pancreatic ATPDase type I.

From the crude cell preparation to the Affigel Blue column, the enzymes of both pancreatic and aorta sources were purified to at least a 1600 fold level (see Tables 1 and 3. After the non-denaturing gel, the quantity of proteins falls under the detection level of the method used, which renders difficult the calculation of a specific activity. However, one can roughly estimate the process to reach about a 10 thousand fold purification, as judged by the density of the ATPDase reaction band relative to other proteins on the non-denaturing electrophoretic gel.

Referring to Table 1, the lectin-binding step is not considered properly as an essential step of the

purification process. This step has been added to show that the aorta ATPDase is a glycoprotein which, when deglycosylated, shifts from a molecular weight of 78 KDa to a molecular of 56 KDa (representing the proteic backbone). Since the lectin-binding step does not achieve the obtention of a pure protein, the most convenient way to obtain a pure protein is to submit the crude cell preparation sequentially to the ion exchange chromatography, the Affigel Blue chromatography and to non-denaturing gel electrophoresis. The identity of the protein is then confirmed by ATP-labelling with FSBA.

Example 3

Partial amino acid sequences

CNBr digests have been obtained from the purified bovine aorta and porcine pancreatic ATPDases. The sequences of the digests are as follows:

Bovine aorta ATPDase:

SEQ.ID. NO.:

Glu Thr Pro Val Tyr Leu Gly Ala Thr Ala Gly
5 10 3

Leu Leu Arg Met Glu
5 4

Ala Asp Lys Ile Leu Ala Asn Xaa Val Ala
5 10

Ser Ser Ile 5

Tyr Pro Phe Asp Phe Gln Gly Ala Arg Ile
5 10 6

Porcine pancreatic ATPDase:

Lys Ser Asp Thr Gln Glu Thr Tyr Gly Ala
5 10

Leu Asp Leu Gly Gly Ala Ser Thr Gln Val
15 20 7

When compared to the sequence which accession number is G2345 (CD39 gene product; Maliszenski et al. 1994), the above partial sequences show a very high degree of homology. The following differences are however found with the CD39 sequence:

In the porcine pancreatic enzyme, Gln^{202} is changed to Lys, the Asn^{204} is changed to Asp, Asn^{205} is changed to Thr.

In the bovine aortic enzyme, Arg^{147} is changed to Lys, Val^{148} is changed to Ile, Asp^{150} is changed to Ala, Gln^{153} is changed to Ala, Arg^{154} is changed to Ser, and Leu^{156} is changed to Ile.

The human CD39 has a predicted molecular weight of 57 KDa, while the apparent molecular of this protein is 78KDa on SDS-PAGE.

Both ATPDases type I and II share a high degree of homology with CD39 for the compared sequenced fragments. CD39 appears to be a human enzyme corresponding to the bovine aortic ATPDase. It is worthwhile noting that the first N-terminal 200 amino acids of CD39 are absent from

the ATPDase type I (pancreatic enzyme). This suggests that the active site of ATPDases is located between the residues 200-510 of CD39 and that part of CD39 is sufficient to provide this activity. It is further worthwhile noting that exact correspondence between the two ATPDases of this invention and the already described ATPDases cannot be established. The human placenta ATPDase (Christoforidis et al. 1995) has a molecular weight of 82KDa while CD39 (also of human origin) has a molecular weight of 78KDa. Due to the differences found in diverse tissues of the same species, extrapolation cannot be done to the effect that the bovine aorta enzyme of this invention is one of the already described enzymes. The obtained partial amino acid sequences indeed already shown differences of sequences which may affect some of the physico-chemical properties of the claimed enzymes when compared to their human counterparts (some of the above-observed substitutions are not conservative ones; the net charge of the enzymes may not be the same and the substituted amino acids may

change the behaviour of the enzymes (optimum pH, sensitivity towards inhibitors, etc ...).

Cross-reactivity between ATPDases I and II:

Antibodies were produced in rabbits against the following amino acid sequence which is common to ATPDase I and CD39:

SEQ. ID. NO.:

Lys Ser Asp Thr Gln Glu Thr Tyr Gly Ala
5 10

Leu Asp Leu Gly Gly Ala
15 8

Figure 5 shows that these antibodies reacted positively with a 78KDa protein present in endothelial extracts of human sources. They also reacted with a protein of 78 KDa of a bovine aorta extract (data not shown). This is an indication that ATPDases I and II share homology of sequence, and that the latter comprises the peptidic sequence of SEQ. ID. No.: 8 or a variant thereof.

A type I ATPDase appears to be present in low amounts in endothelial cells as shown by the detection of a faint band corresponding to this protein (54KDa) in Figure 5.

CONCLUSIONS:

-Considering that the ATPDase has an antihemostatic role in the saliva of blood-feeding insects and leeches (Rigbi et al., 1987);

-considering that Côté et al. (1992) have demonstrated bovine ATPDase type II has platelet anti-aggregant properties by converting ADP to AMP;

-considering the low Km of the aorta type II enzyme (μM), the optimum pH of catalysis pH 7.5-8.0, its localization at the surface of endothelial and smooth muscle cells of blood vessels (Côté et al., 1992);

-considering that the purified enzyme keeps its original characteristics;

it sounds predictable that the aorta enzyme produced in the present invention can be introduced in

the circulatory system of mammalians to reduce platelet aggregation and thrombogenicity.

Furthermore, considering that a crude microsomal bovine ATPDase type II fraction has been successfully conjugated to agarose and that the conjugate could reduce ADP induced platelet aggregation (Hirota et al., 1987);

-considering that a semi-purified plant ATPDase has been successfully coupled to the following matrices: CM-cellulose, copolymers of L-alanine and L-glutamic acid, polyaspartic acid, polygalacturonic acid, Elvacite 2008™ (methyl methacrylate) and ethylene-maleic acid co-polymer (Patel et al., 1969);

we propose that the purified ATPDase type II can be coupled to artificial polymers/biomaterials to reduce thrombogenicity (platelet aggregation).

Therefore, pharmaceutical compositions for use in the reduction of platelet aggregation and thrombogenicity are under the scope of the invention. These compositions should contain, as an active

ingredient, the ATPDase type II of this invention combined to an acceptable carrier without excluding any form or formulation of such compositions. Finally, considering that the sequenced CD39 appears to correspond to a human counterpart of the bovine ATPDase type II enzyme of this invention, the use of CD39 or variants or a part thereof for reducing platelet aggregation and thrombogenicity is also part of this invention.

A new process for producing an ATPDase comprising the steps of:

- obtaining a host which comprises a nucleic acid encoding a protein having the amino acid sequence defined in SEQ. ID. NO.: 1, or a variant thereof, or a part thereof, said variant or part being capable of converting ATP to ADP and ADP to AMP;

- culturing said host in a culture medium supporting the growth of said host and the expression of said nucleic acid;

- recovering the ATP diphosphohydrolase from the culture medium or from said host; and

- purifying the ATP diphosphohydrolase

is also part of the invention. Preferably the nucleic acid is the one defined in SEQ ID NO.: 2, or a part or a variant thereof, which part or variant is capable of producing an ATP diphosphohydrolase.

The present invention has been described hereinabove; it will become apparent to the skilled reader that variations could be brought thereto without departing from the teachings of the present disclosure. Such variations are under the scope of this invention.

TABLE 3

ATPDase purification

Results of one out of three preparations is presented. Determinations were carried out in triplicate.

* Laliberté *et al.* showed a 160 fold purification for the ZGM as compared to the homogenate using ADP as the substrate.

Steps	Total protein	Total activity	Specific activity (ATP)	Yield	Purification factor	Hydrolysis rates ATP/ADP
	mg	*units*	*units/mg*	*%*	*fold*	
ZGM	20.0	60.8	3.0	-	(160)*	1.3
ZGM + Triton X-100	20.0	40.6	2.0	100	1	1.3
100,000 g supernatant of solubilized ZGM	17.6	37.0	2.1	91	1.1	1.3
DEAE column	3.5	28.8	8.3	71	4.2	1.3
Affi-Gel blue column	0.31	13.8	45	34	23	1.3

BIBLIOGRAPHY:

Baykov et al. (1988). Anal. Biochem. 171: 266-270.

Bradford, M (1976). Anal. Biochem. 72: 248-254.

Côté et al. (1991). BBA 1078: 187-191.

Côté et al. (1992). BBA 1139: 133-142.

Christoforidis et al. (1995), Eur. J. Biochem. 234: 66-74.

Hirota et al. (1987). Thrombosis Res. 45: 201-209.

Laemmli (1970). Nature 227: 680-685.

Lebel et al. (1980). J. Biol. Chem. 255: 1227-1233.

Maliszenski et al. (1994). J. Immunol. : 3574-3583

Patel et al. (1969). BBA 178: 626-629.

Rigbi et al. (1987). Comp. Biochem. Physiol. 87B: 567-573.

Yagi et al. (1989). Eur. J. Biochem. 180: 509-513.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANTS: BEAUDOIN, Adrien R.
SÉVIGNY, Jean

(ii) TITLE OF THE INVENTION: ATP-DIPHOSPHOHYDROLASES, PROCESS OF PURIFICATION THEREOF AND PROCESS OF PRODUCING THEREOF BY RECOMBINANT TECHNOLOGY

(iii) NUMBER OF SEQUENCES: 8

(iv) CORRESPONDENCE ADDRESS:

(A) ADDRESSEE: GOUDREAU GAGE DUBUC & MARTINEAU WALKER
(B) STREET: 800 PLACE VICTORIA, SUITE 3400
(C) CITY: MONTREAL
(D) STATE: QUEBEC
(E) COUNTRY: CANADA
(F) ZIP: H4Z 1E9

(v) COMPUTER READABLE :

(A) MEDIUM TYPE: DISKETTE 1.44
(B) COMPUTER: IBM Compatible
(C) OPERATING SYSTEM: WINDOWS 3.1
(D) SOFTWARE: WORDPERFECT 6.1

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER:
(B) FILING DATE: APRIL 10, 1996
(C) CLASSIFICATION:

(vii) PRIOR APPLICATION DATA:

(A) APPLICATION: U.S.S.N 08/419,204
(B) FILING DATE: APRIL 10, 1995

(viii) ATTORNEY/AGENT INFORMATION:

(A) NAME: Jean H. DUBUC, Gaétan PRINCE, Alain M. LECLERC
(C) REFERENCE/DOCKET NUMBER: DH/10857.146

(ix) TELECOMMUNICATION INFORMATION:

(A) TELEPHONE: (514) 397-4335
(B) TELEFAX: (514) 397-4382

(2) INFORMATION FOR SEQ ID NO: 1

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 510 AMINO ACIDS
(B) TYPE: AMINO ACID
(C) STRANDEDNESS: SINGLE
(D) TOPOLOGY: LINEAR

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1

```
Met Glu Asp Thr Lys Glu Ser Asn Val Lys Thr Phe Cys Ser Lys
 5                  10                  15
Asn Ile Leu Ala Ile Leu Gly Phe Ser Ser Ile Ile Ala Val Ile
 20                  25                  30
Ala Leu Leu Ala Val Gly Leu Thr Gln Asn Lys Ala Leu Pro Glu
 35                  40                  45
Asn Val Lys Tyr Gly Ile Val Leu Asp Ala Gly Ser Ser His Thr
 50                  55                  60
Ser Leu Tyr Ile Tyr Lys Trp Pro Ala Glu Lys Glu Asn Asp Thr
 65                  70                  75
Gly Val Val His Gln Val Glu Glu Cys Arg Val Lys Gly Pro Gly
 80                  85                  90
Ile Ser Lys Phe Val Gln Lys Val Asn Glu Ile Gly Ile Tyr Leu
 95                 100                 105
Thr Asp Cys Met Glu Arg Ala Arg Glu Val Ile Pro Arg Ser Gln
110                 115                 120
His Gln Glu Thr Pro Val Tyr Leu Gly Ala Thr Ala Gly Met Arg
125                 130                 135
Leu Leu Arg Met Glu Ser Glu Glu Leu Ala Asp Arg Val Leu Asp
140                 145                 150
Val Val Glu Arg Ser Leu Ser Asn Tyr Pro Phe Asp Phe Gln Gly
155                 160                 165
```

```
Ala Arg Ile Ile Thr Gly Gln Glu Glu Gly Ala Tyr Gly Trp Ile
                170                 175                 180
Thr Ile Asn Tyr Leu Leu Gly Lys Phe Ser Gln Lys Thr Arg Trp
                185                 190                 195
Phe Ser Ile Val Pro Tyr Glu Thr Asn Asn Gln Glu Thr Phe Gly
                200                 205                 210
Ala Leu Asp Leu Gly Gly Ala Ser Thr Gln Val Thr Phe Val Pro
                215                 220                 225
Gln Asn Gln Thr Ile Glu Ser Pro Asp Asn Ala Leu Gln Phe Arg
                230                 235                 240
Leu Tyr Gly Lys Asp Tyr Asn Val Tyr Thr His Ser Phe Leu Cys
                245                 250                 255
Tyr Gly Lys Asp Gln Ala Leu Trp Gln Lys Leu Ala Lys Asp Ile
                260                 265                 270
Gln Val Ala Ser Asn Glu Ile Leu Arg Asp Pro Cys Phe His Pro
                275                 280                 285
Gly Tyr Lys Lys Val Val Asn Val Ser Asp Leu Tyr Lys Thr Pro
                290                 295                 300
Cys Thr Lys Arg Phe Glu Met Thr Leu Pro Phe Gln Gln Phe Glu
                305                 310                 315
Ile Gln Gly Ile Gly Asn Tyr Gln Gln Cys His Gln Ser Ile Leu
                320                 325                 330
Glu Leu Phe Asn Thr Ser Tyr Cys Pro Tyr Ser Gln Cys Ala Phe
                335                 340                 345
Asn Gly Ile Phe Leu Pro Pro Leu Gln Gly Asp Phe Gly Ala Phe
                350                 355                 360
Ser Ala Phe Tyr Phe Val Met Lys Phe Leu Asn Leu Thr Ser Glu
                365                 370                 375
Lys Val Ser Gln Glu Lys Val Thr Glu Met Met Lys Lys Phe Cys
                380                 385                 390
Ala Gln Pro Trp Glu Glu Ile Lys Thr Ser Tyr Ala Gly Val Lys
                395                 400                 405
Glu Lys Tyr Leu Ser Glu Tyr Cys Phe Ser Gly Thr Tyr Ile Leu
                410                 415                 420
Ser Leu Leu Leu Gln Gly Tyr His Phe Thr Ala Asp Ser Trp Glu
                425                 430                 435
His Ile His Phe Ile Gly Lys Ile Gln Gly Ser Asp Ala Gly Trp
                440                 445                 450
Thr Leu Gly Tyr Met Leu Asn Leu Thr Asn Met Ile Pro Ala Glu
                455                 460                 465
Gln Pro Leu Ser Thr Pro Leu Ser His Ser Thr Tyr Val Phe Leu
                470                 475                 480
```

Met Val Leu Phe Ser Leu Val Leu Phe Thr Val Ala Ile Ile Gly
485 490 495
Leu Leu Ile Phe His Lys Pro Ser Tyr Phe Trp Lys Asp Met Val
500 505 510

(2) INFORMATION FOR SEQ ID NO: 2

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1818 BASE PAIRS
(B) TYPE: NUCLEIC ACID
(C) STRANDEDNESS: SINGLE
(D) TOPOLOGY: LINEAR

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2

ACCACACCAA GCAGCGGCTG GGGGGGGGAA AGACGAGGAA AGAGGAGGAA 50
AACAAAAGCT GCTACTTATG GAAGATACAA AGGAGTCTAA CGTGAAGACA 100
TTTTGCTCCA AGAATATCCT AGCCATCCTT GGCTTCTCCT CTATCATAGC 150
TGTGATAGCT TTGCTTGCTG TGGGGTTGAC CCAGAACAAA GCATTGCCAG 200
AAAACGTTAA GTATGGGATT GTGCTGGATG CGGGTTCTTC TCACACAAGT 250
TTATACATCT ATAAGTGGCC AGCAGAAAAG GAGAATGACA CAGGCGTGGT 300
GCATCAAGTA GAAGAATGCA GGGTTAAAGG TCCTGGAATC TCAAAATTTG 350
TTCAGAAAGT AAATGAAATA GGCATTTACC TGACTGATTG CATGGAAAGA 400
GCTAGGGAAG TGATTCCAAG GTCCCAGCAC CAAGAGACAC CCGTTTACCT 450
GGGAGCCACG GCAGGCATGC GGTTGCTCAG GATGGAAAGT GAAGAGTTGG 500
CAGACAGGGT TCTGGATGTG GTGGAGAGGA GCCTCAGCAA CTACCCCTTT 550
GACTTCCAGG GTGCCAGGAT CATTACTGGC CAAGAGGAAG GTGCCTATGG 600
CTGGATTACT ATCAACTATC TGCTGGGCAA ATTCAGTCAG AAAACAAGGT 650
GGTTCAGCAT AGTCCCATAT GAAACCAATA ATCAGGAAAC CTTTGGAGCT 700
TTGGACCTTG GGGGAGCCTC TACACAAGTC ACTTTTGTAC CCCAAAACCA 750
GACTATCGAG TCCCCAGATA ATGCTCTGCA ATTTCGCCTC TATGGCAAGG 800
ACTACAATGT CTACACACAT AGCTTCTTGT GCTATGGGAA GGATCAGGCA 850
CTCTGGCAGA AACTGGCCAA GGACATTCAG GTTGCAAGTA ATGAAATTCT 900
CAGGGACCCA TGCTTTCATC CTGGATATAA GAAGGTAGTG AACGTAAGTG 950
ACCTTTACAA GACCCCCTGC ACCAAGAGAT TTGAGATGAC TCTTCCATTC 1000
CAGCAGTTTG AAATCCAGGG TATTGGAAAC TATCAACAAT GCCATCAAAG 1050
CATCCTGGAG CTCTTCAACA CCAGTTACTG CCCTTACTCC CAGTGTGCCT 1100
TCAATGGGAT TTTCTTGCCA CCACTCCAGG GGGATTTTGG GGCATTTTCA 1150
GCTTTTTACT TTGTGATGAA GTTTTTAAAC TTGACATCAG AGAAAGTCTC 1200
TCAGGAAAAG GTGACTGAGA TGATGAAAAA GTTCTGTGCT CAGCCTTGGG 1250
AGGAGATAAA AACATCTTAC GCTGGAGTAA AGGAGAAGTA CCTGAGTGAA 1300

```
TACTGCTTTT CTGGTACCTA CATTCTCTCC CTCCTTCTGC AAGGCTATCA        1350
TTTCACAGCT GATTCCTGGG AGCACATCCA TTTCATTGGC AAGATCCAGG        1400
GCAGCGACGC CGGCTGGACT TTGGGCTACA TGCTGAACCT GACCAACATG        1450
ATCCCAGCTG AGCAACCATT GTCCACACCT CTCTCCCACT CCACCTATGT        1500
CTTCCTCATG GTTCTATTCT CCCTGGTCCT TTTCACAGTG GCCATCATAG        1550
GCTTGCTTAT CTTTCACAAG CCTTCATATT TCTGGAAAGA TATGGTATAG        1600
CAAAAGCAGC TGAAATATGC TGGCTGGAGT GAGGAAAAAA TCGTCCAGGG        1650
AGCATTTTCC TCCATCGCAG TGTTCAAGGC CATCCTTCCC TGTCTGCCAG        1700
GGCCAGTCTT GACGAGTGTG AAGCTTCCTT GGCTTTTACT GAAGCCTTTC        1750
TTTTGGAGGT ATTCAATATC CTTTGCCTCA AGGACTTCGG CAGATACTGT        1800
CTCTTTCATG AGTTTTTC                                           1818
```

(2) INFORMATION FOR SEQ ID NO: 3

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 11 AMINO ACIDS
(B) TYPE: AMINO ACID
(C) STRANDEDNESS: SINGLE
(D) TOPOLOGY: LINEAR

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3

```
Glu Thr Pro Val Tyr Leu Gly Ala Thr Ala Gly
 5                  10
```

(2) INFORMATION FOR SEQ ID NO: 4

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 5 AMINO ACIDS
(B) TYPE: AMINO ACID
(C) STRANDEDNESS: SINGLE
(D) TOPOLOGY: LINEAR

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4

```
Leu Leu Arg Met Glu
 5
```

(2) INFORMATION FOR SEQ ID NO: 5

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 13 AMINO ACIDS

(B) TYPE: AMINO ACID
(C) STRANDEDNESS: SINGLE
(D) TOPOLOGY: LINEAR

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5

Ala Asp Lys Ile Leu Ala Asn Xaa Val Ala Ser Ser Ile
5 10

(2) INFORMATION FOR SEQ ID NO: 6

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 10 AMINO ACIDS
(B) TYPE: AMINO ACID
(C) STRANDEDNESS: SINGLE
(D) TOPOLOGY: LINEAR

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6

Tyr Pro Phe Asp Phe Gln Gly Ala Arg Ile
5 10

(2) INFORMATION FOR SEQ ID NO: 7

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 AMINO ACIDS
(B) TYPE: AMINO ACID
(C) STRANDEDNESS: SINGLE
(D) TOPOLOGY: LINEAR

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7

Lys Ser Asp Thr Gln Glu Thr Tyr Gly Ala Leu Asp Leu Gly Gly
5 10 15
Ala Ser Thr Gln Val
20

(2) INFORMATION FOR SEQ ID NO: 8

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 16 AMINO ACIDS
(B) TYPE: AMINO ACID

(C) STRANDEDNESS: SINGLE
(D) TOPOLOGY: LINEAR

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8

Lys Ser Asp Thr Gln Glu Thr Tyr Gly Ala Leu Asp Leu Gly Gly
5 10 15
Ala

WHAT IS CLAIMED IS:

1. An isolated and purified ATP diphosphohydrolase obtainable from bovine aorta characterized by the following physico-chemical properties:

-a catalytic unit of a molecular weight on denaturing polyacrylamide gel electrophoresis of about 78 KDa in its native form;

-a deglycosylated form of said catalytic unit of a molecular weight on SDS-PAGE of about 56 KDa; and characterized in that it comprises the amino acid sequences defined in SEQ. ID. NOs. 3 to 6.

2. An ATP diphosphohydrolase as defined in claim 1 further comprising the amino acid sequence defined in SEQ. ID. No.: 8 or a variant thereof.

3. An isolated and purified ATP diphosphohydrolase obtainable from pig pancreatic zymogen granules

characterized by the following physico-chemical properties:

-a catalytic unit of a molecular weight on denaturing polyacrylamide gel electrophoresis of about 54 KDa in its native form;

-a deglycosylated form of said catalytic unit of a molecular weight on SDS-PAGE of about 35 KDa; and characterized in that it comprises the amino acid sequence defined in SEQ. ID. NO.: 7.

4. A process for purifying an ATP-diphosphohydrolase enzyme from a tissue capable to convert ATP to ADP and ADP to AMP which comprises:

a) obtaining a sub-cellular microsomal fraction from an homogenate of said tissue;

b) solubilizing said microsomal fraction in the presence of a non-ionic detergent;

c) centrifuging said solubilized microsomal fraction to obtain a supernatant containing said enzyme;

d) submitting said supernatant to an ion-exchange chromatography to obtain a first enzyme eluate;

e) submitting said first eluate to an affinity column chromatography to obtain a second enzyme eluate; and

f) submitting said second eluate to a separation step on a non-denaturing gel electrophoresis to recover said enzyme free of any contaminant, the presence of said contaminant being monitored by overstaining said gel in a silver nitrate dye or Coomassie Blue dye.

5. A process according to claim 4 wherein said ion exchange chromatography is achieved on a column containing Diethylaminoethyl (DEAE).

6. A process according to claim 5 wherein said column is a DEAE agarose column.

7. A process according to claim 4 or 5 wherein said affinity column chromatography is achieved on an Affigel™ Blue column.

8. A process according to claim 4, 5, 6 or 7 wherein said non-ionic detergent is Triton X-100™.

9. A process according to claim 4, 5, 6, 7 or 8 wherein an aliquot of said enzyme is further submitted after step f) to a polyacrylamide gel electrophoresis under denaturing conditions to verify its homogeneity and to obtain its apparent molecular weight.

10. A process according to claim 9 wherein said enzyme is obtained from pig pancreatic zymogen granules and has an apparent molecular weight of 54 Kilodaltons.

11. A process according to claim 9 wherein said enzyme is obtained from bovine aortic intima layer and has an apparent molecular weight of about 78 Kilodaltons.

12. A process according to claim 10 wherein, between steps e) and f), a step of deglycosylation is included, and whereby the apparent molecular weight is shifted from 54 to 35 KDa.

13. A process according to claim 11 herein, between steps e) and f), a step of deglycosylation is included, and whereby the apparent molecular weight is shifted from 78 to 56 KDa.

14. The use of the ATP diphosphohydrolase of claim 1 or 2, for reducing platelet aggregation and thrombogenicity.

15. The use of an ATP diphosphohydrolase for reducing platelet aggregation and thrombogenicity, said ATP diphosphohydrolase having the amino acid sequence defined in SEQ. ID. NO.: 1, or a variant thereof, or a part thereof, said variant or part being capable of converting ATP to ADP and ADP to AMP.

16. A composition for use in the reduction of platelet aggregation and thrombogenicity which comprises as an active ingredient the ATP diphosphohydrolase of claim 1 or 2 or an ATP diphosphohydrolase which sequence is defined in SEQ. ID. NO.: 1, or a variant or a part thereof, which variant or part has an ATP diphosphohydrolase activity, in an acceptable pharmaceutical carrier.

17. A process for producing an ATP diphosphohydrolase which comprises the steps of:

- obtaining a host which comprises a nucleic acid encoding a protein having the amino acid sequence defined in SEQ. ID. NO.: 1, or a variant thereof, or a part thereof, said variant or part being capable of converting ATP to ADP and ADP to AMP;

- culturing said host in a culture medium supporting the growth of said host and the expression of said nucleic acid;

- recovering the ATP diphosphohydrolase from the culture medium or from said host; and

- purifying the ATP diphosphohydrolase.

18. A process as defined in claim 17, wherein said nucleic acid has a sequence defined in SEQ. ID. NO.: 2, a variant thereof or a part thereof, said variant or part being capable of producing an ATP diphosphohydrolase which converts ATP to ADP and ADP to AMD.

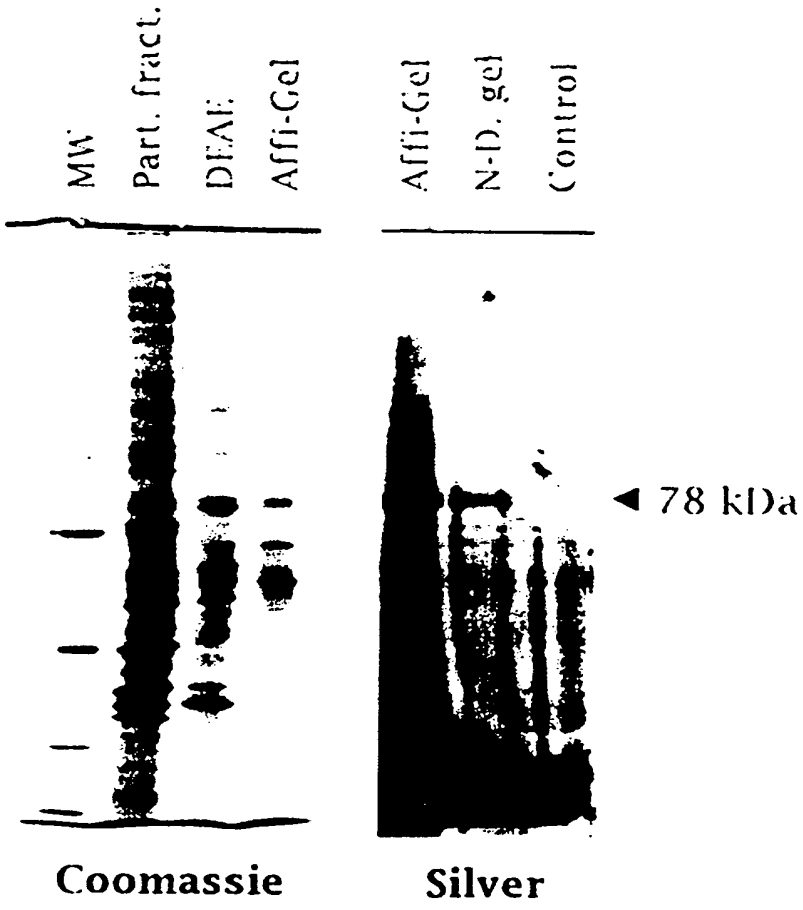

MW
Part. fract.
DEAE
Affi-Gel
Affi-Gel
N-D. gel
Control
78 kDa
Coomassie
Silver

Fig. 1

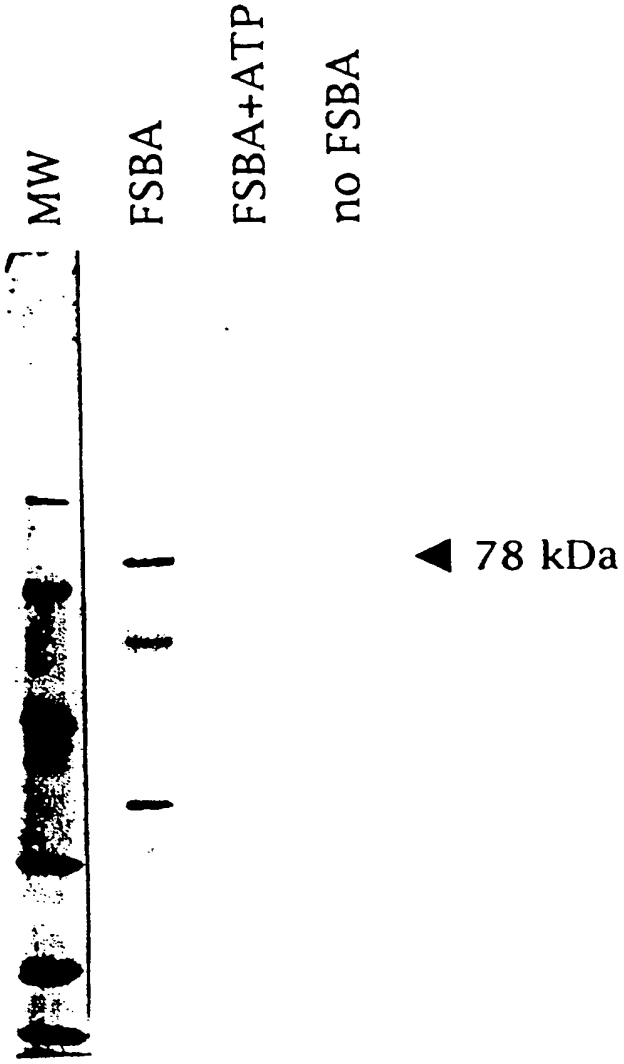
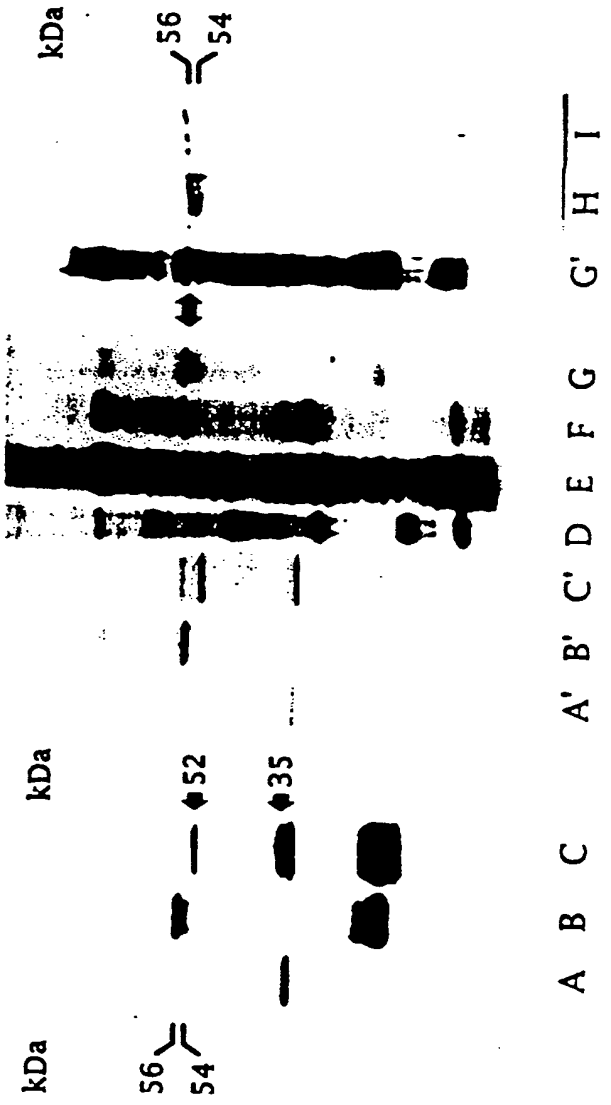


Fig. 2

Fig. 3



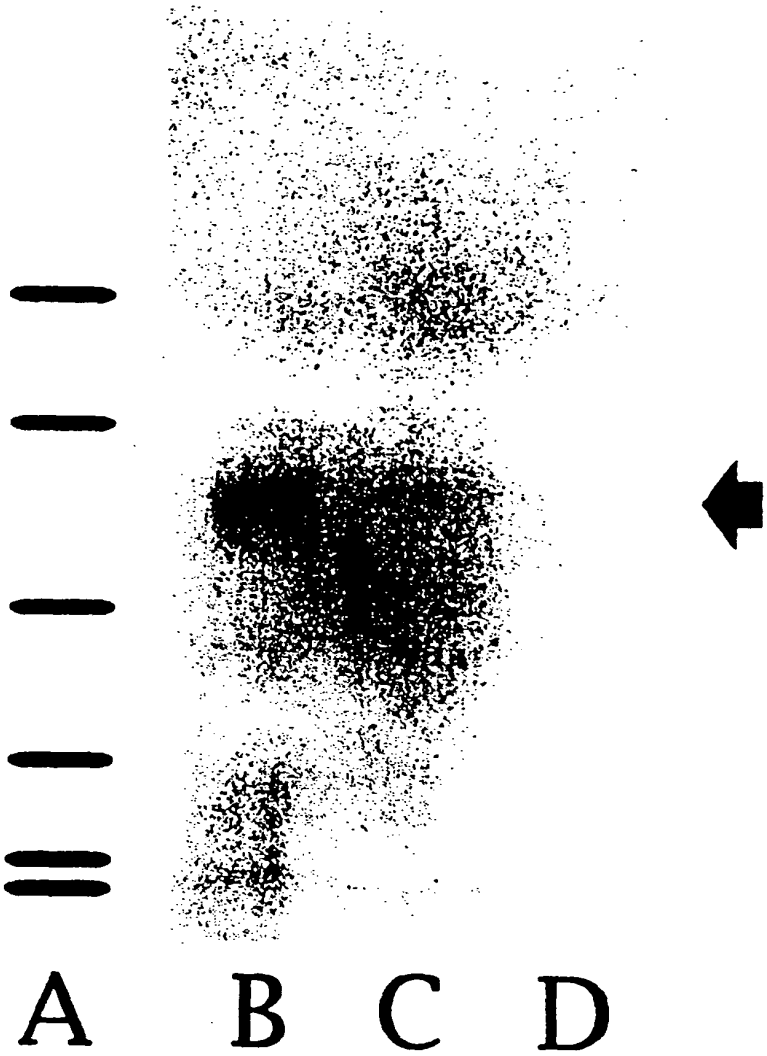
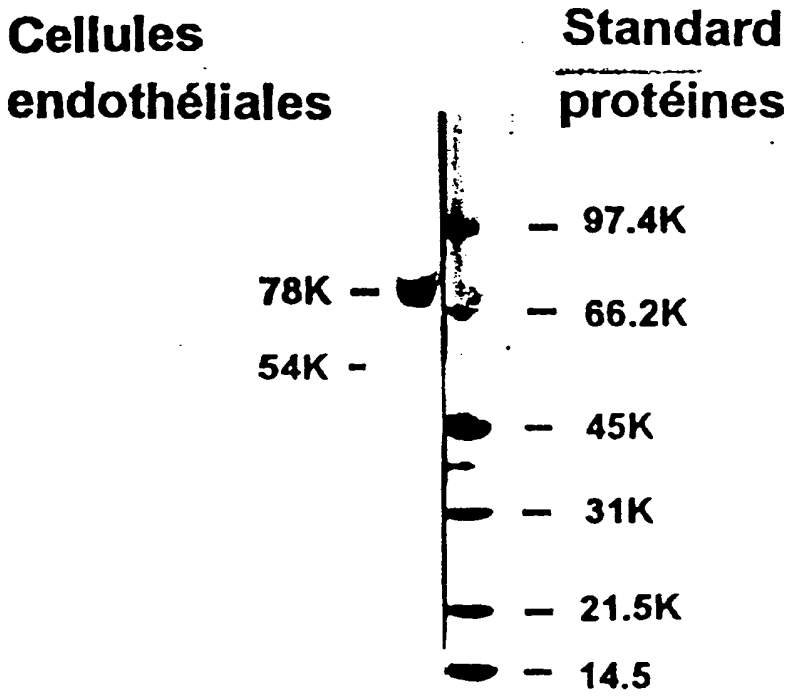


Fig. 4



Western Blot

Fig. 5